

[illegible]

1X buffer 0	10mM Tris-HCl pH 7.5 at 37°C, 10mM MgCl ₂ and 0.1mg/ml BSA.
1X buffer c	10mM Tris-HCl pH 7.5 at 37°C, 10mM MgCl ₂ , 50mM NaCl and 0.1mg/ml BSA.
1X buffer d	50mM Tris-HCl pH 7.5 at 37°C, 10mM MgCl ₂ , 10mM NaCl and 0.1mg/ml BSA.
1X buffer R	10mM Tris-HCl pH 8.5 at 37°C, 10mM MgCl ₂ , 100mM KCl and 0.1mg/ml BSA.
Buffer Y/Tango	
1X	33mM Tris-acetate pH 7.9 at 37°C, 10mM Mg-acetate, 66mM K-acetate and 0.1mg/ml BSA.
2X	66mM Tris-acetate pH 7.9 at 37°C, 20mM Mg-acetate, 132mM K-acetate and 0.2mg/ml BSA.

How do I do a Double Digest?

1. Select each enzyme of interest.
2. If the optimal coo-cool buffer for both enzymes is the same, use that buffer.
3. If not, use the buffer in which the two enzymes are most active.
4. If neither buffer is suitable for both REs, refer to the column headed "**Y+Tang**" buffer concentration for "double digestion".

If the same Y+Tang:buffer concentration is recommended for both enzymes, use it.

If both 1X and 2X are suitable, choose 2X.

If different Y+Tang:buffer concentrations are recommended, perform the first digest at 1X final concentration, then add extra 10X Y+Tang:buffer (0.125 ml) to the reaction mixture to bring the final buffer concentration to 2X to perform the second digest.

If Y+Tang:buffer is "not recommended" (star activity appears at a 5-fold overdigestion (50x $\times$ 1 hr)), perform sequential digests in RE optimal buffers.

****** indicates the appearance of star activity at a 10-fold overdigestion (10 units $\times$ 1 hr). To avoid this, use a smaller excess of RE.

