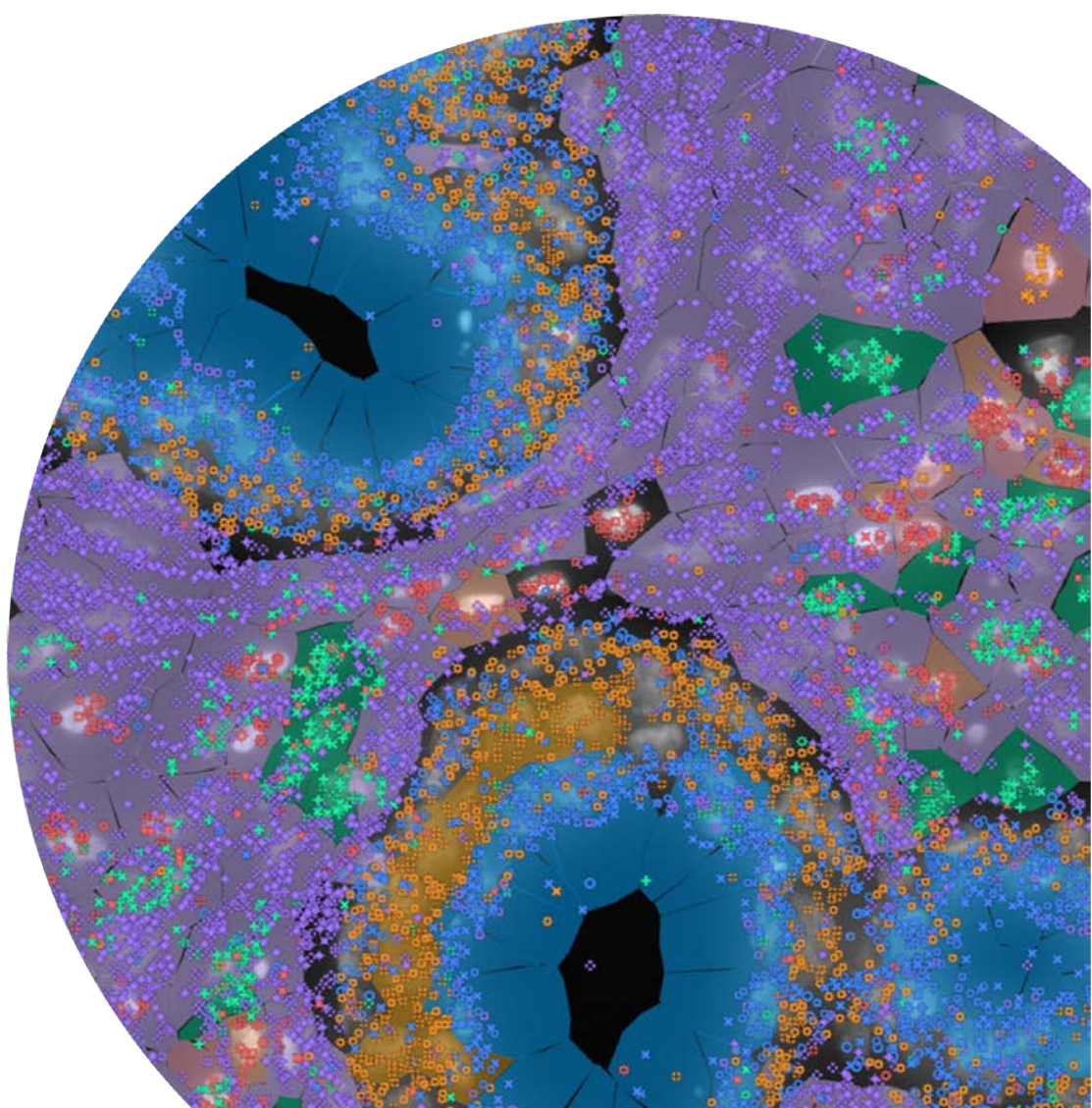


# *In Situ Analyzer 10x Genomics* Xenium Analysis

# Report



# Project Information

Client Name	Samples
Company/Institution	Samples
Order Number	Order Number
Species	Species
Number of Slide	-
Number of FOV	-
Region Area( $\mu\text{m}^2$ )	-
Total Cell Area( $\mu\text{m}^2$ )	-
Panel Used	Panel ID
Number of Target Genes(RNA)	-

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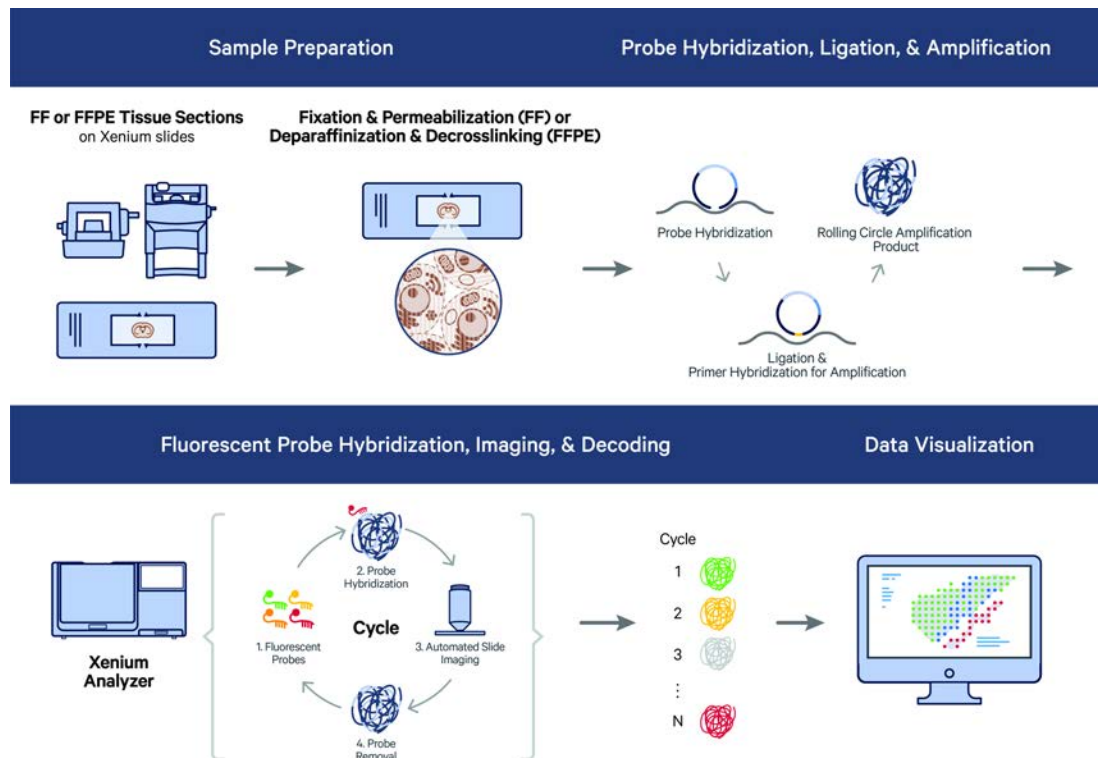
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# 1. Xenium Technology

## 1.1. Overall Workflow



## 1.2. Xenium In Situ Slide

- The Xenium Slide is designed to analyze mRNA in tissue sections derived from formalin fixed & paraffin embedded (FFPE) tissue samples.
- Tissue Slides and Xenium Slides are loaded into their Xenium instrument, where they are brought into proximity with one another.
- Each Xenium Slide has a 12 × 24 mm imageable area on which tissue sections can be placed, allowing multiple tissue pieces to be included on each slide. Two Xenium slides can be analyzed in each Xenium Analyzer run. Xenium slides have been designed for optimal assay performance and to minimize tissue detachment across multiple sample types.
- Xenium slides have been designed for optimal assay performance and to minimize tissue detachment across multiple sample types.
- 400s of RNA targets alongside multiplexed protein.

## 1. 3. Xenium Analyzer

- The input data for the Xenium Analyzer are Xenium slides containing tissue sections.
- The Xenium Analyzer captures vertical stacks of images every cycle and in every channel for multiple fields of view. The stacks are processed and stitched together to build a single image of the tissue section.

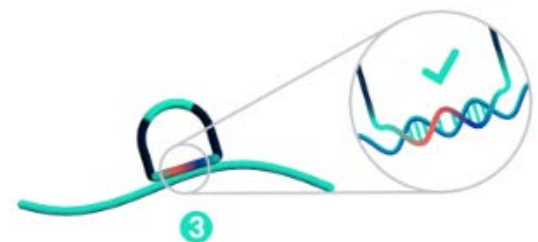
### Steps 1 & 2: Dual target recognition & hybridization

Both probe arms must stably hybridize to their target. If only one arm hybridizes, the probe is unstable and will be washed off in the post hybridization wash. The padlock probes also contain a gene-specific barcode sequence that is used to generate a unique optical signature of each transcript.



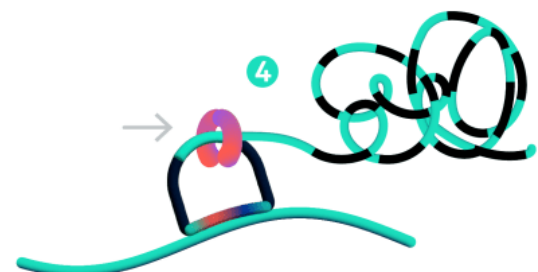
### Step 3: Probe ligation

Only when both probe arms are stably hybridized to the target with a perfect match does proximal probe ligation occur. Partially bound or mis-matching probes are not ligated and cannot be amplified.



### Step 4: Rolling circle amplification (RCA)

Only ligated probes are amplified, enriching for true target recognition events and suppressing off-target events. The RCA approach creates a robust and strong signal allowing single probe detection.



## 2. Summary of Data Production

### 2.1. Data Statistics

#### 2.1.1. Key Metrics

Slide Name	FOV	Region Area ( $\mu\text{m}^2$ )	Total Cell Area ( $\mu\text{m}^2$ )	Number of cells detected	Decoded transcripts per $100\mu\text{m}^2$
Slide_1	1	-	-	-	-
Slide_1	2	-	-	-	-
Slide_2	1	-	-	-	-
Slide_2	2	-	-	-	-

- FOV : Field of view
- Median transcript per cell : The median number of transcripts per cell. Cells with zero transcripts are excluded from the calculation.
- Decoded transcripts per  $100\mu\text{m}^2$  : Counts the number of high-quality decoded transcripts and divides it by the total estimated tissue area to get a transcript density.

#### 2.1.2. Decoding Yield

Slide Name	FOV	Percent of all gene transcripts that are high quality	Total high quality decoded transcripts
Slide_1	1	-	-
Slide_1	2	-	-
Slide_2	1	-	-
Slide_2	2	-	-

- Percent of all gene transcripts that are high quality : The percent of transcripts from all genes on the gene panel that decode with high quality ( $\geq Q20$ ).
- Total high quality decoded transcripts : The total number of decoded gene transcripts that decode with high quality ( $\geq Q20$ ).

#### 2.1.3. Segmentation Metrics

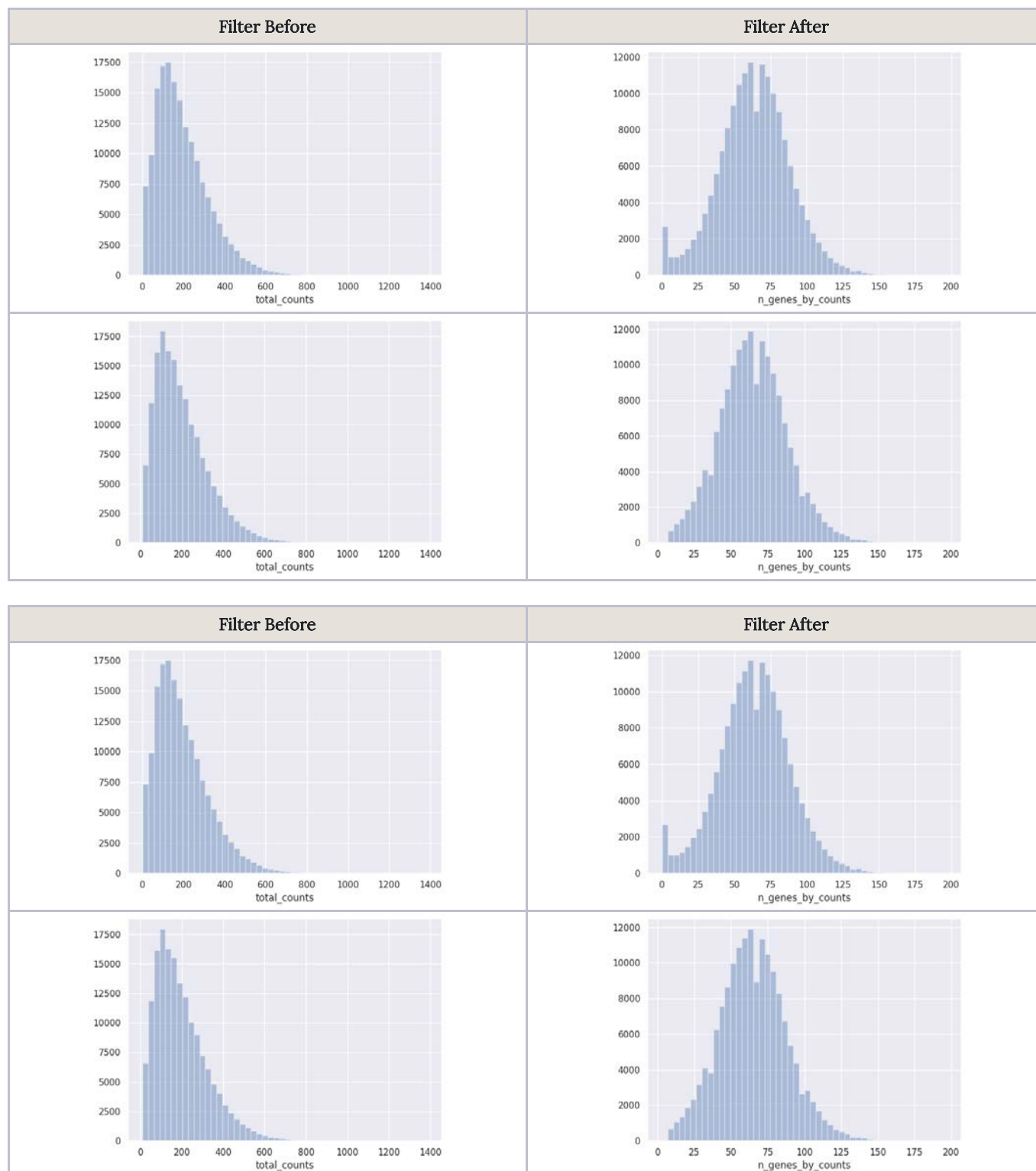
Slide Name	FOV	Fraction of empty cells	Percent of transcripts within cells	Cells per $100\mu\text{m}^2$	Median genes per cell	Median transcripts per cell
Slide_1	1	-	-	-	-	-
Slide_1	2	-	-	-	-	-
Slide_2	1	-	-	-	-	-
Slide_2	2	-	-	-	-	-

- Fraction of empty cells : The percentage of all cells without decoded high-quality transcripts.
- Percent of transcripts within cells : Percent of high-quality transcripts that are found within cells.
- Cells per  $100\mu\text{m}^2$  : The density of cells per 100 microns squared.
- Median genes per cell : The median number of unique genes detected per cell. Cells with zero transcripts are excluded from the calculation.
- Median transcripts per cell : The median number of transcripts per cell. Cells with zero transcripts are excluded from the calculation.



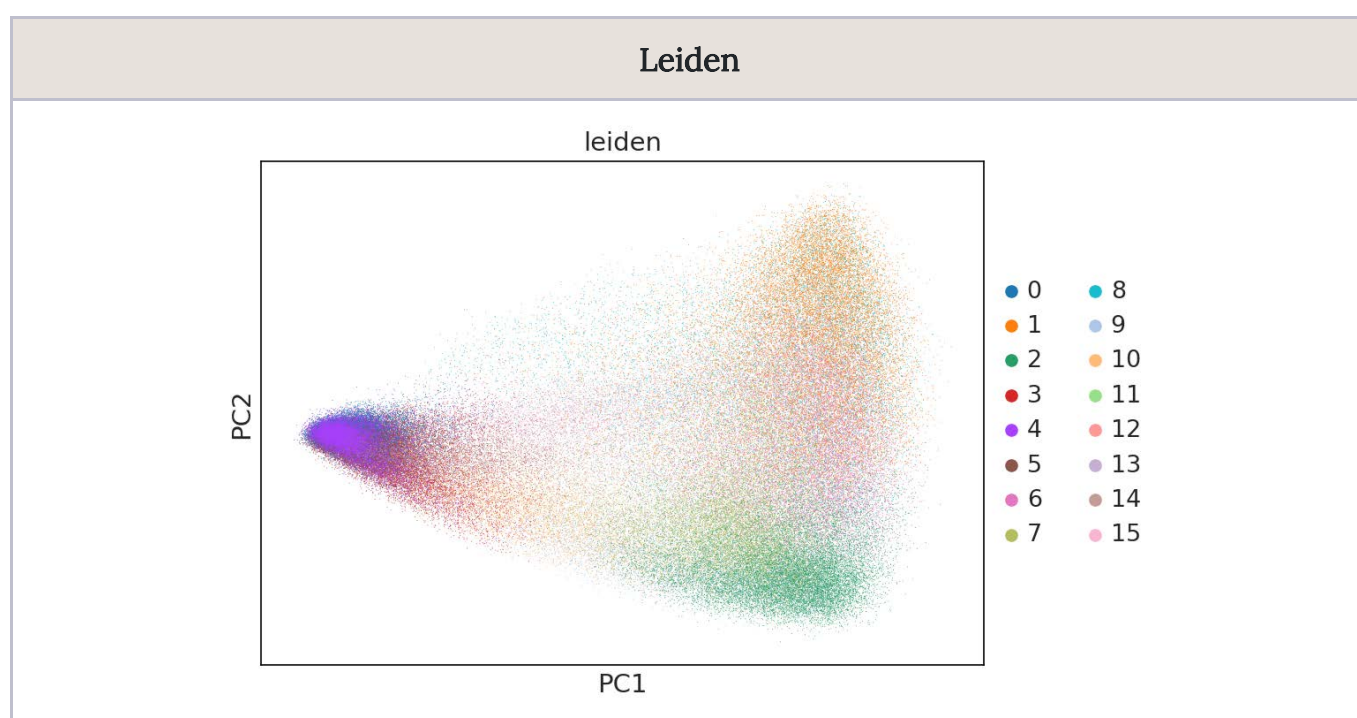
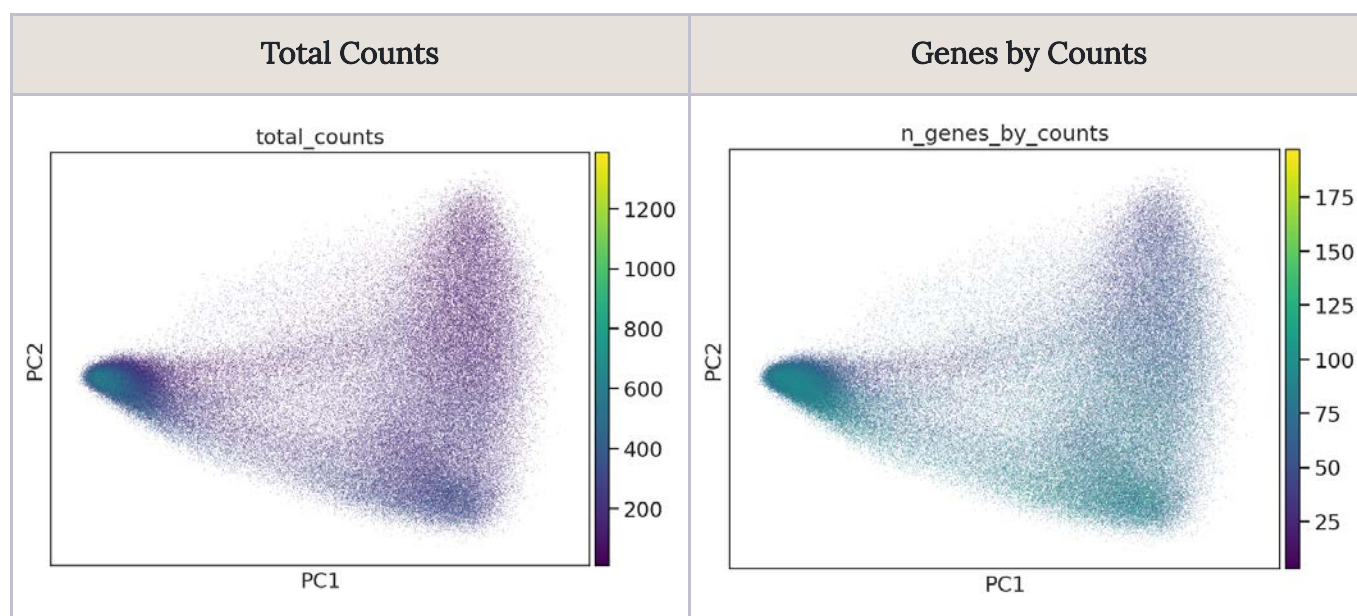
## 3. Result of analysis transcripts per cells

### 3.1. Stat QC plot



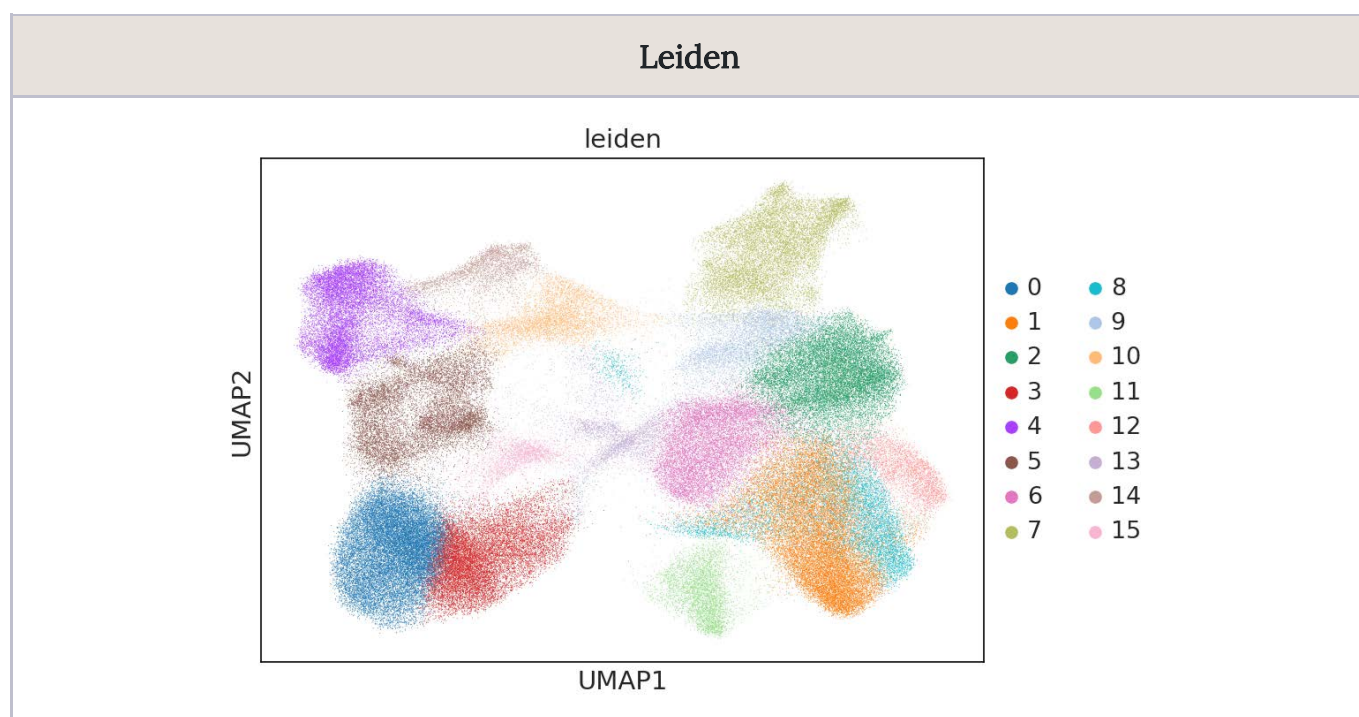
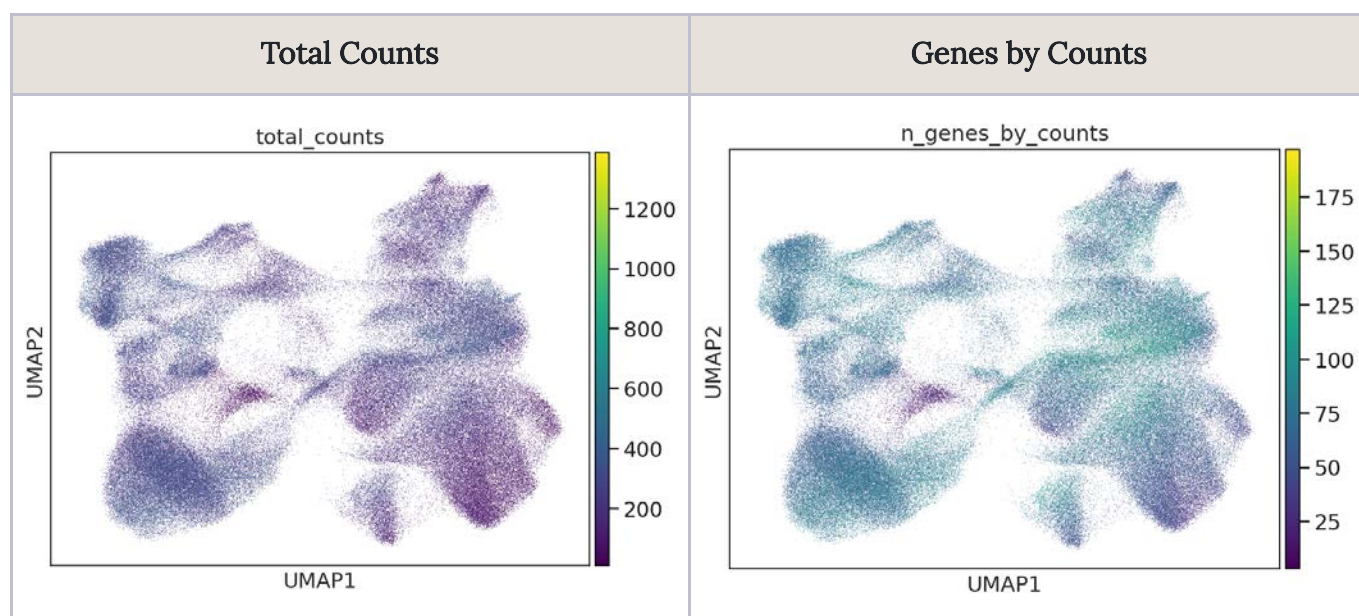
- Total counts : Total transcripts per cell.
- N genes by counts : Unique transcripts per cell.
- Default threshold value : Minimum transcripts per cell is 10, minimum unique transcripts per cell is 5.

## 3. 2. PCA plot

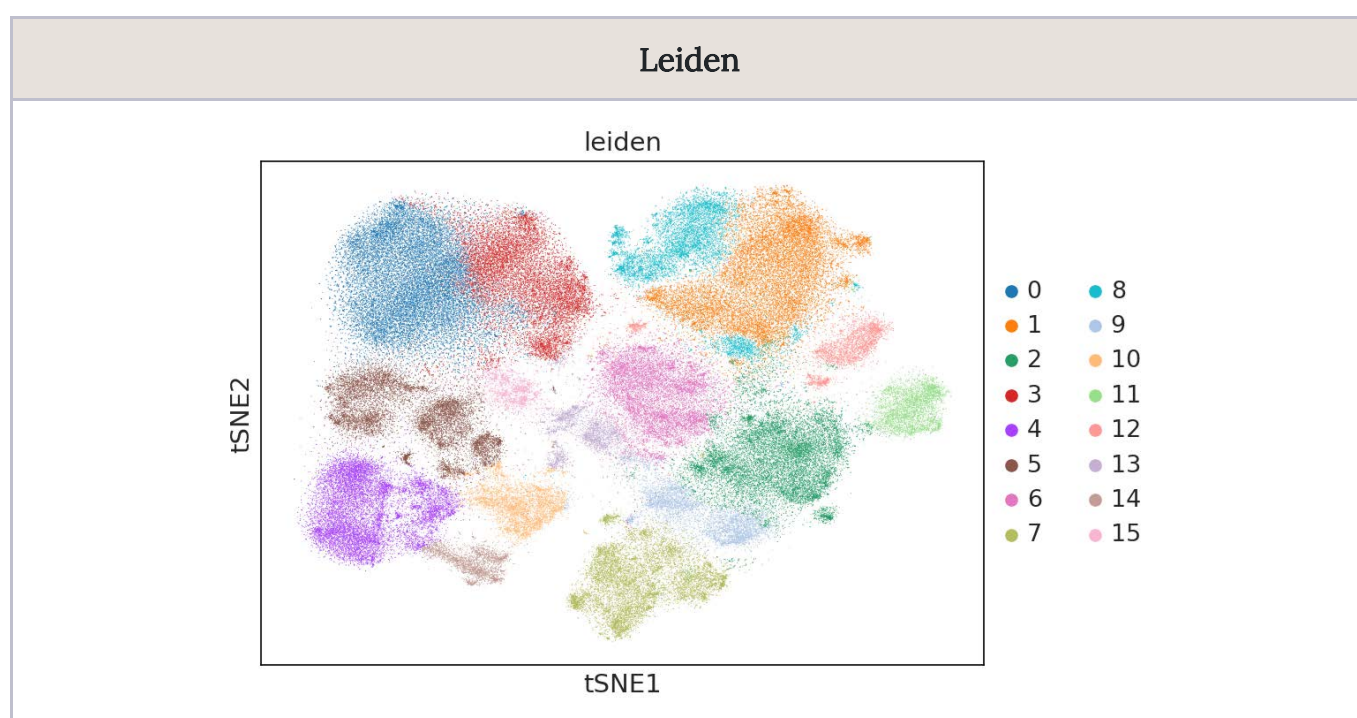
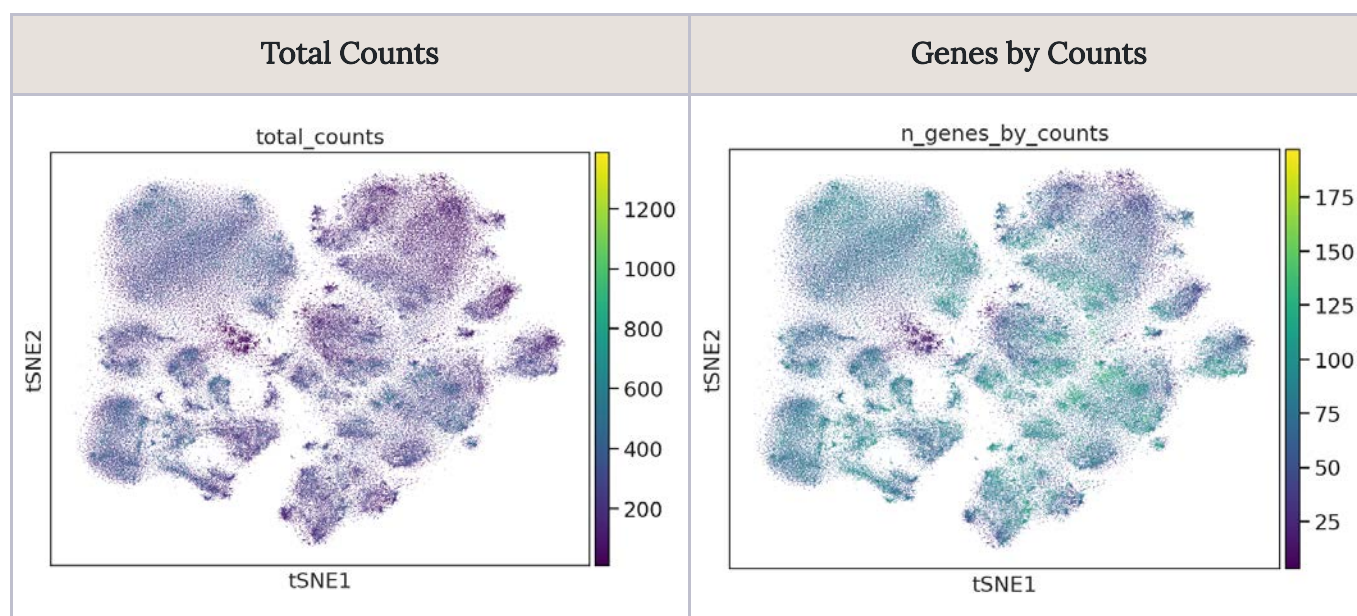




### 3. 3. UMAP plot

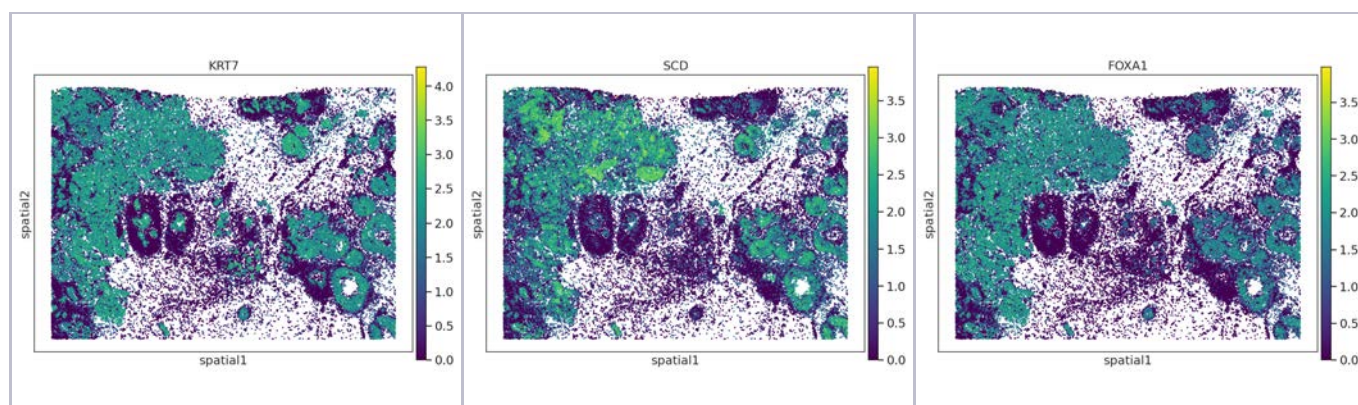


### 3. 4. t-SNE plot

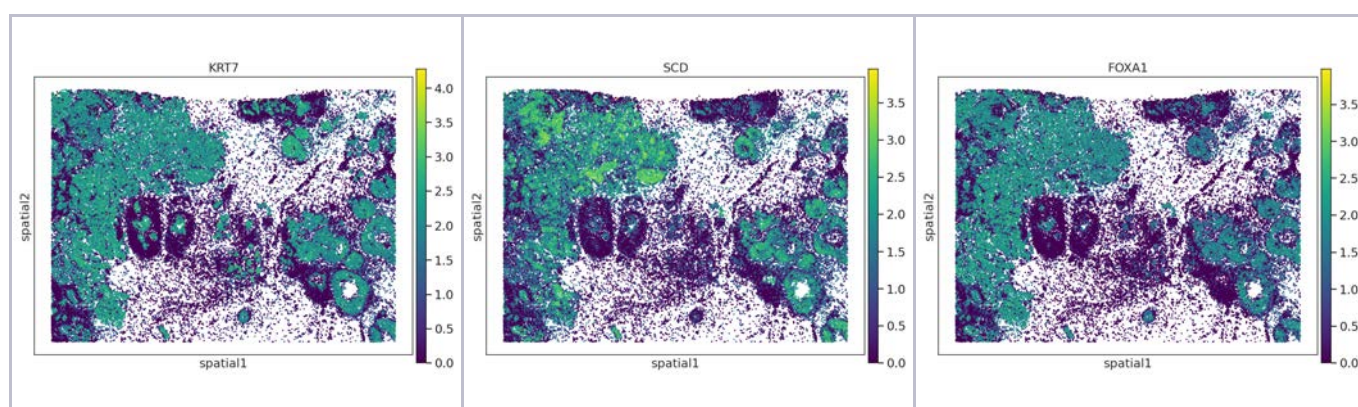


### 3. 5. Moran's I score TOP3

- FOV1



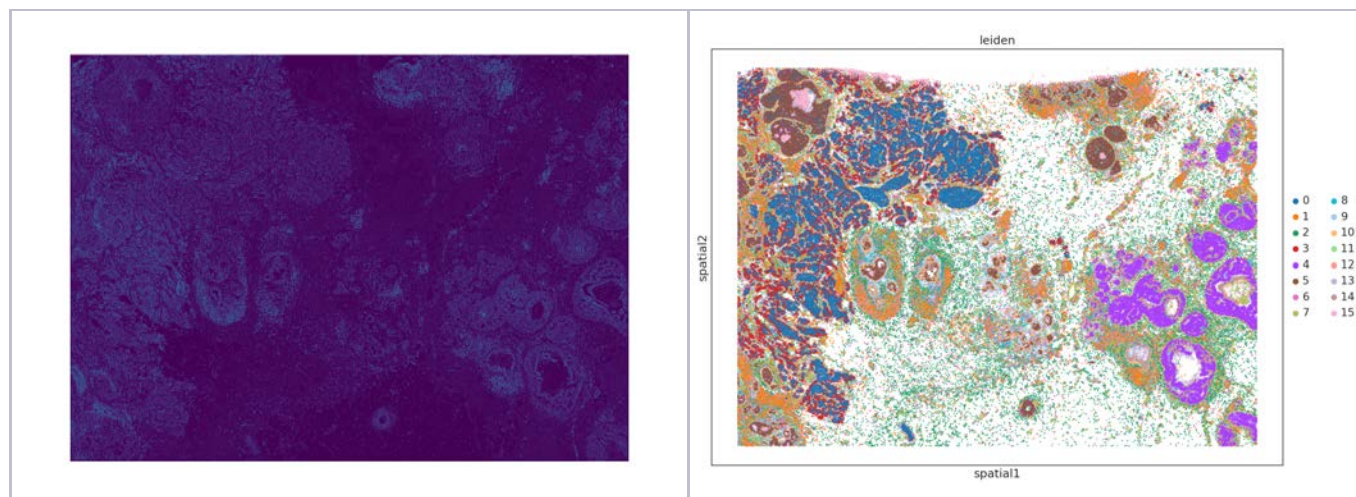
- FOV2



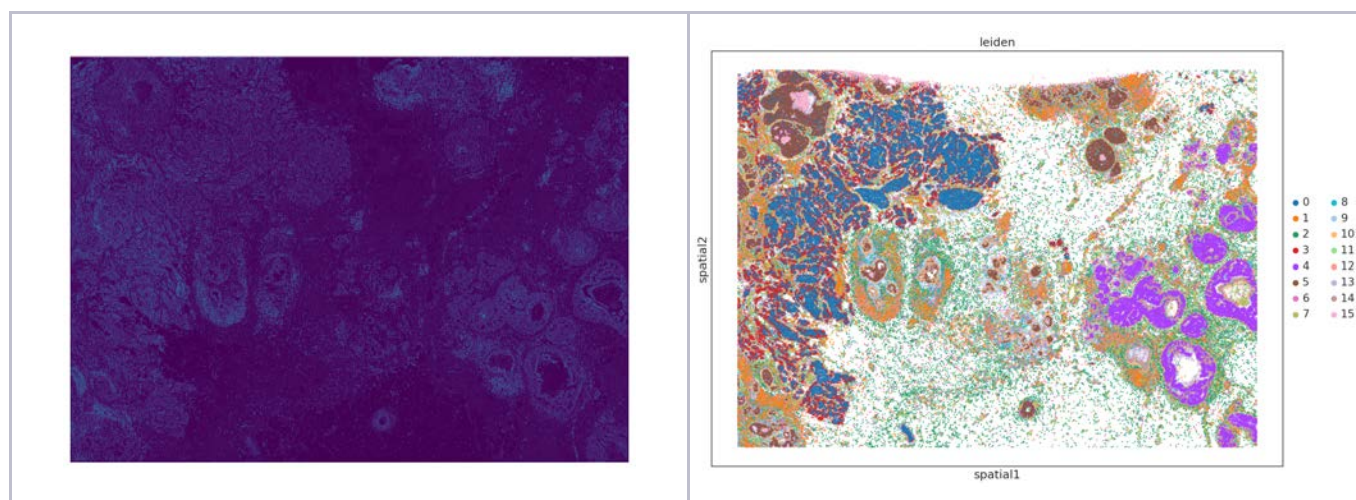


## 3. 6. Spatial Scatter Plot

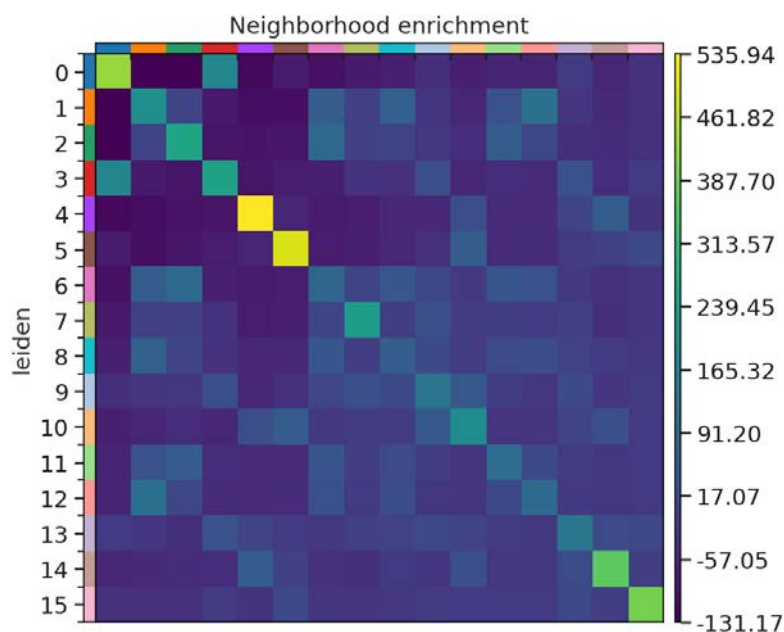
- FOV1



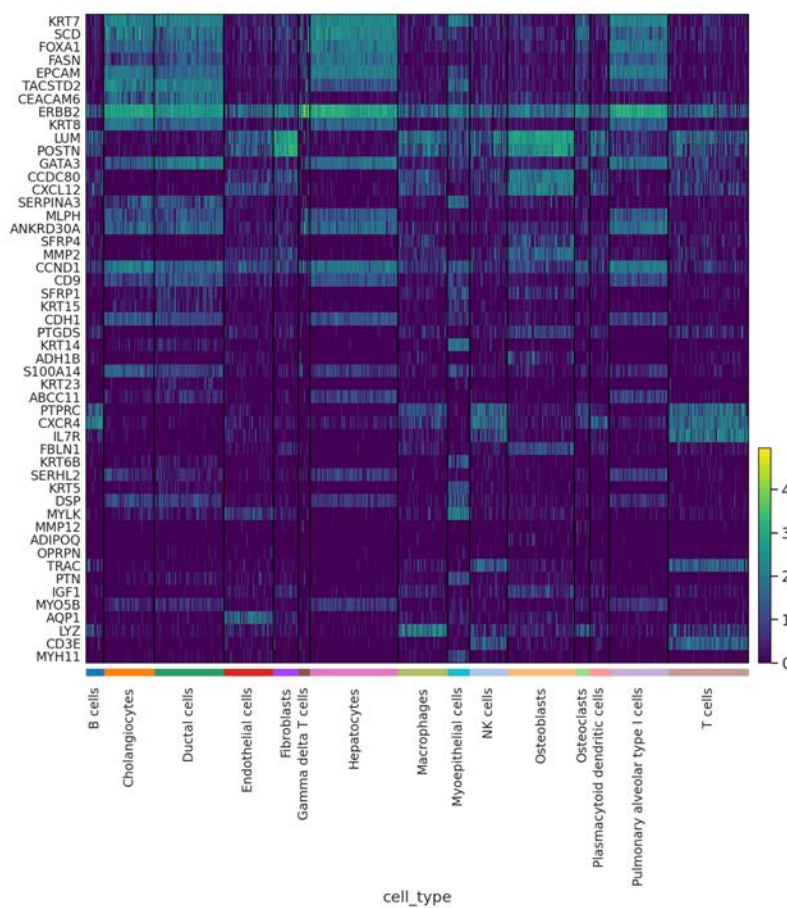
- FOV2



## 3. 7. Neighborhood Enrichment



## 3. 8. Gene X Cell Heatmap



## 4. Data Download Information

### 4. 1. Analysis Results

Download link	File size
Result.tar (md5sum : )	File Sizes

### 4. 2. Folder Structure

```

Xenium Result
├── [Slide]
├── analysis
│   ├── clustering
│   │   ├── kmeans [2~10] cluster — cluster.csv
│   │   └── graphclust — cluster.csv
│   ├── diffexp
│   │   ├── kmeans [2~10] cluster — differential expression.csv
│   │   └── graphclust — differential expression.csv
│   ├── pca
│   │   └── gene expression 10 components — variance.csv components.csv dispersion.csv projection.csv features selected.csv
│   └── umap
│       └── gene expression 2 components — projection.csv
├── cell feature matrix — matrix.mtx.gz barcodes.tsv.gz features.tsv.gz
├── aux outputs
│   └── fov_locations.json
├── cell feature plot
│   ├── [before|after]_[total_counts|n_genes_by_counts].png
│   ├── [pca|umap|tsne]_[total_counts|n_genes_by_counts|leiden|cell_type].png
│   ├── [heatmap|spatial_scatter|neighborhood_enrichment]_[leiden|cell_type].png
│   ├── moran_I_score_[1st|2nd|3rd].png
│   └── morphology_focus.ome.png
├── analysis_summary.html
├── analysis.zarr.zip
├── cell_boundaries.csv.gz
├── cell_boundaries.parquet
├── cell_feature_matrix.h5
├── cell_feature_matrix.zarr.zip
├── cells.csv.gz
├── cells.parquet
├── cells.zarr.zip
├── experiment.xenium
├── gene_panel.json
├── metrics_summary.csv
├── morphology_focus.ome.tif
├── morphology_mip.ome.tif
├── morphology.ome.tif
├── nucleus_boundaries.csv.gz
├── nucleus_boundaries.parquet
├── transcripts.csv.gz
├── transcripts.parquet
└── transcripts.zarr.zip

```



## 5. Appendix

### 5.1. Literature References

scanpy	<a href="https://scanpy.readthedocs.io/en/stable/index.html">https://scanpy.readthedocs.io/en/stable/index.html</a>
squidpy	<a href="https://squidpy.readthedocs.io/en/stable/index.html">https://squidpy.readthedocs.io/en/stable/index.html</a>
decoupler	<a href="https://decoupler-py.readthedocs.io/en/latest/index.html">https://decoupler-py.readthedocs.io/en/latest/index.html</a>
panglaoDB	<a href="https://panglaoDB.se/#google_vignette">https://panglaoDB.se/#google_vignette</a>
moran's I	<a href="https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9745188/">https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9745188/</a>

### 5.2. Result Appendix

- **Clustering** : with graph-based and K-means results. Graph-based clustering (under graphclust) is run once as it does not require a pre-specified number of clusters. K-means (under kmeans) is run for  $K=2..N$  where K corresponds to the number clusters, and N=10 by default. Each value of K has its own results directory.
- **Principal Component Analysis (PCA)** : which contains a total of five files listing the features used in the dimension reduction i.e., to reduce the feature space. These results are used to perform clustering.
- **UMAP** : contains the Uniform Manifold Approximation and Projection results.
- **t-SNE** : is a technique that visualizes high dimensional data by giving each point a location in a two or three-dimensional map.
- **Moran's I Score** : measures spatial autocorrelation using feature locations and feature values simultaneously. The spatial autocorrelation tool utilizes a multidimensional and multi-directional factors. The Moran's I index will be a value between -1 and 1. Positive spatial autocorrelation will show values that are clustered. Negative autocorrelation is dispersed. Random is close to zero.

## 5. 2. Xenium Gene Expression Panel

Gene List
Gene List