

IHC Detect Kit for Mouse Primary Antibody Data Sheet

Cat.NO.: PK10010

General Information:

Sample type: FFPE tissues

Assay type: Immunohistochemistry

Secondary antibody host species: Goat

Secondary antibody species specificity: Mouse

Description:

The PK10010 kit is designed to facilitate signal detection during IHC analysis of FFPE tissue samples using primary antibodies raised in mice. The kit consists of Polymer HRP-Goat anti-Mouse Secondary Antibody and an ultra-sensitive Chromogen for highly sensitive detection of mouse primary antibodies. The kit also contains an optimized primary antibody dilution buffer that preserves primary antibody activity and helps lower non-specific background staining.

Note:

* If antigen retrieval buffer, counter staining reagent and mounting media are needed, please see our IHC Prep & Detect Kit for Mouse Primary Antibody (Cat.NO. PK10018).

Components:

Part NO.	Component	Size
HC000	Quenching Buffer (RTU)	6 mL
HC004	Primary Antibody Diluent (RTU)	24 mL
HC005M	Polymer HRP-Goat anti-Mouse Secondary Antibody	6 mL
HC006	Chromogen Component A	0.4 mL
HC007	Chromogen Component B	8 mL

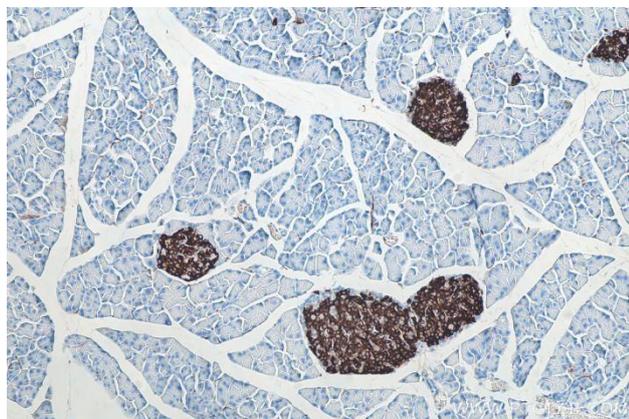
Size: 60 tests (may vary depending on tissue size)

Storage: All the reagents are stored at 2-8°C. The kit is stable for 6 months from the date of receipt.

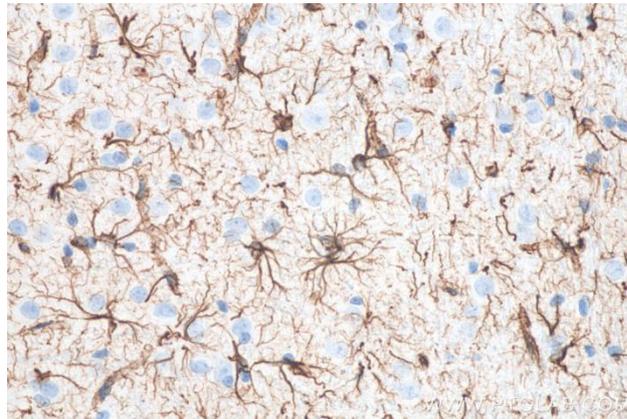
Application Note:

This kit is recommended for FFPE tissues and not for frozen sections.

Validation Data:



IHC analysis of mouse pancreas tissue using Proteintech's Insulin mouse monoclonal antibody (66198-1-Ig) and IHC Detect Kit for Mouse Primary Antibody (PK10010).



IHC analysis of rat brain tissue using Proteintech's GFAP mouse monoclonal antibody (60190-1-Ig) and IHC Detect Kit for Mouse Primary Antibody (PK10010).

Recommended Protocol:

1. Prepare slides from tissues sections following routine methods. Deparaffinize tissues with xylene and re-hydrate using a decreasing ethanol gradient.
2. Perform antigen retrieval following the manufacturer's recommendations of the primary antibody.
3. Quenching (optional): Based on our experience, endogenous peroxidase quenching is not necessary in most cases and can sometimes even have adverse effects on staining. However, we have included a quenching reagent in this kit since the quenching step is widely recommended in publications.
 - 3.1 Wash the slides with 1x wash buffer. Drain off the liquid on the slides and absorb any residual buffer around the tissue section (making the slides as dry as possible). Use a hydrophobic IHC pen to draw a circle on the glass slides around tissue sections (optional).

- 3.2 Apply 30-80 μ L (depending on the size of the tissue) of the ready-to-use Quenching Buffer (Part NO. HC000) to the slide, covering the entire tissue. Leave the slides at room temperature for 10 minutes. Briefly wash the slides with 1x wash buffer after quenching.
4. Block the tissues with blocking buffer (e.g., 3% BSA in PBS) for 30 minutes at room temperature. Drain the liquid off the slides and absorb the blocking buffer around the tissue section.
* The Primary Antibody Diluent (Part NO. HC004) could also be used as alternative of blocking buffer.
 5. Use the Primary Antibody Diluent (Part NO. HC004) in the kit to dilute your primary antibody (ONLY primary antibodies raised in mice should be used with this kit). Apply 80-200 μ L (depending on the size of the tissue) of the diluted primary antibody to the slide, covering the entire tissue. Incubate at room temperature in a wet box with a lid for 1 hour.
 6. Wash the slides three times with 1x wash buffer following primary antibody incubation. Drain the liquid off the slides and absorb any residual buffer around the tissue section. Apply 30-80 μ L (depending on the size of the tissue) of the ready-to-use Polymer HRP-Goat anti-Mouse Secondary Antibody (Part NO. HC005M) to the slide, covering the entire tissue. Incubate at room temperature in a wet box with a lid for 30 minutes.
 7. Wash the slides three times with 1x wash buffer following secondary antibody incubation. Drain the liquid off the slides and absorb any residual buffer around the tissue section. Use pipette to prepare an appropriate volume of Chromogen in a microcentrifuge tube. Ratio of solutions Chromogen Component A (Part NO. HC006) to Chromogen Component B (Part NO. HC007) is 1:20.
* Depending on the size of the tissue sample, approximately 30-80 μ L/slide is needed. Use pipette to add enough Chromogen to cover the tissue. Lay slides flat on the benchtop at room temperature for 5 minutes, or until a brown color develops. If color is visible, stop reaction early by washing with 1x wash buffer.
 8. Perform counter staining with hematoxylin.
 9. Wash the slides with wash buffer or pure water. Dehydrate tissues using an increasing ethanol gradient followed by xylene. Mount the slides with cover slides and analyze with a light microscope once they are fully dried.